THE HOLOTHURINOGENINS-II¹ METHOXYLATED NEO-HOLOTHURINOGENINS

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(Received in USA 27 Augusr I%8 ; *Received in the UK for pubIicuiion 3 December 1968)*

Abstract-Holothurin A, on hydrolysis in methanol-HCl (0-2N; 50°) yielded the methoxylated neoholothurinogenins: 128-methoxy-7,8-dihydro-22,25-oxidoholothurinogenin (III), its desoxy analog (IV), **12B_methoxy-7,8-diiydr~24,2S-dehydro (V), 12&dimcthoxy-?,8-diiydro (VI), and 12&methoxy-22** hydroxy-7,8-dihydroholothurinogenin (VII). Their structures and relationships to the neo-holothurino**genins, tbe aglycone moieties as they exist in holothurin A, and the holothurinogenins, obtained on strong acid hydrolysis of holothurin A, are discussed. The NMR spectra of the methoxylated neo-holothurin~** genins in benzene, chloroform and pyridine are reported.

MANY species of sea-cucumbers in the family of holothuroidea of the phylum Echinodermata contain a poisonous principle, presumably for their defense against predators. Holothurin, the toxic saponin principle, obtained from the Carbibean species, Actinopyga agassizi, has been the subject of a number of physiological² and chemical investigations.3 Recently, we have reported' on the structures of a few of the holothurinogenins, 22,25-oxidoholothurinogenin (I), its 17-desoxy derivative (II), and holothurinogenin U, the latter a mixture of closely related genins possessing the same annular portion as compounds I or II, but with open side-chains. The holothurinogenins, derived from the glycoside holothurin by strong acid hydrolysis, differ, however, from the corresponding aglycone moieties, neo-holothurinogenins, as they exist in native holothurin. The holothurinogenins contain the 7 :8, 9 : 11 heteroannular conjugated diene system, holothurin does not. This diene system is formed during the strong acid hydrolysis. Additionally, the question has been raised as to whether other substantial changes in the triterpenoid moiety, e.g. closure of the oxide ring, lactone ring rearrangement, had resulted from strong acid treatment.

To obtain the unmodified aglycones we have investigated the mild non-aqueous acid hydrolysis of holothurin $A³$, the principal glycosidic constituent of holothurin. In this paper we report on the methoxylated neo-holothurinogenins,⁴ 12 β -methoxy-7,8-dihydro-22,25-oxidoholothurinogenin (III), its 17 desoxy analog, compound IV; 12B-methoxy-7,8-dihydro-24,25-dehydro (V), 12B, 25-dimethoxy-7,8-dihydro (VI), and 12B-methoxy-22-acetoxy-7,8-dihydroholothurinogenin (VII) (Fig. 1) obtained from holothurin A by methanol-HCl hydrolysis. Their structural relationships to the neo-holothurinogenins, the known holothurinogenins, and NMR spectra are discussed.

Hydrolysis of holothurin A in CH₃OH-HCl($0.2N$; 50 $^{\circ}$) effected the removal of the entire sugar residue without formation of the conjugated diene system and gave

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FIG. 1

a mixture of mono- and dimethoxylated neo-holothurinogenins. Holothurin A contains no OMe group other than the one associated with 3-methoxyglucose³ in its sugar residue, which additionally contains xylose, glucose, and quinovose. The separation of the individual components was achieved by chromatography of their acetates. Their identification (described in detail below), was confirmed by their conversion on strong acid treatment to their corresponding known holothurinogenins. The monomethoxylated compounds III, IV, V and VII gave the known holothurinogenins I, II, VIII⁵ and IX (griseogenin),⁶ respectively (Fig. 1). The dimethoxylated neo-holothurinogenin VI, the principal constituent of the mixture, also gave VIII.

The hydrolysis of dihydroholothurin A in MeOH-HCl was investigated, since the possibility existed that even under these mild conditions the 22,25-oxido ring common to compounds III and IV was formed via cyclixation of a precursor with an hydroxyl at C_{22} and a double bond issuing from C_{25} . The 22,25-oxido compounds III and IV were again obtained, as was compound VII. Neither the dimethoxylated holothurinogenin VI, nor the related unsaturated monomethoxylated holothurinogenin V were found. In their place the dihydromonomethoxylated neo-holothurinogenin X was obtained. Clearly, the second OMe group in compound VI resulted from the addition of methanol to a double bond. A comparison of the NMR spectra (cf. Table 1) of compounds V, VI, VII and X, demonstrated that this OMe group was situated on C_{25} and obviously derived by addition of methanol to its immeduate precursor, compound V.

The fact that each of the methoxylated neo-holothurinogenins gave, on strong acid treatment, the conjugated 7 : 8, 9 : **1 1-diene** system, characteristic of the holothurinogenins, and the ease with which the one OMe common to all these compounds was substituted, suggested that an OH group, α to a double bond, present in the parent holothurin, was replaced. Confirmation was obtained by hydrolysis of dihydroholothurin A in dioxan-HCl. The mixture of mono-unsaturated aglycones (as acetates) exhibited a broad singlet at δ 5.6 ppm (not present in the methoxylated compounds), which may be attributed to the proton on the C atom bearing the replaceable OH group, since by subsequent treatment of this mixture with methanol-HCl this singlet was eliminated with the concomitant appearance of a broad singlet at ca. δ 3.5, characteristic of the proton on the C atom bearing the aforementioned OMe group (Table 1). The NMR spectra and spin decoupling experiments (discussed below) of the various methoxylated precursors, were in accord with the assigned structures and established both the position, C_{12} , and stereochemistry, β , of the OMe group in each of the aforementioned precursors. The IR spectra of the precursors, especially with respect to the position of the lactone absorption, corresponded closely to that observed for their derived holothurinogenins.

These findings strongly support the presumption that, except for the replacement of an OH on C_{12} by a OMe, the methoxylated neoholothurinogenins correspond in structure to the aglycone moieties as they exist in holothurin A. The question as to the stereochemistry, α , and/or β , of the C₁₂-OH in holothurin A is discussed in the following paper. We discuss below the identification of compounds III, IV, V, VI and VII.

12 β -Methoxy-7,8-dihydro-22,25-oxidoholothurinogenin-3-acetate (III). $C_{33}H_{50}O_7$, m.p. 273°, $\lceil \alpha \rceil_0^{25}$ – 60°. The compound exhibited no significant absorption in the UV above 215 mµ. The IR (CS₂) spectrum showed the presence of an OH (3570), γ -lactone (1766), acetyl (1730), and side-chain ether (1124 cm⁻¹). These bands correspond to those observed¹ for 22,25-oxodoholothurinogeninacetate (I). The seventh oxygen was accounted for in the OMe group (1089 cm⁻¹). The NMR spectra (60 Mc/s, $CDCl₃$) showed the presence of one vinyl hydrogen (5.37, broad singlet); and two hydrogens (overlapping) centered at 4.12 (C_{12} —H and C_{22} —H). In benzene and pyridine a clear separation into a triplet and a broad singlet was achieved. The triplet, centered at 4.08 ($J = 7$ c/s) and 4.26 ($J = 7$ c/s) in benzene and pyridine, respectively, corresponded in both chemical shift and coupling constant to the observed for the C_{22} —H proton in 22,25-oxidoholothurinogenin (I). The second signal, which appeared as a broad singlet at 4.37 and 4.50 in benzene and pyridine, respectively, may be ascribed to the proton on the C atom bearing the OMe group. Its disposition, α to the double bond, was indicated by its relative downfield chemical shift as compared to such protons in saturated molecules.' This conclusion was confirmed by spin decoupling experiments at 100 Mc/s. Irradiation (CDCl₃) at 4.12 resulted in narrowing of the broad singlet (5.37) of the vinylic hydrogen. Of the various possibilities for the position of the double bond, $5:6,7:8,9:11$, with the OMe group at C₇, C₆ or C₁₂, respectively, the following considerations clearly establish β sub-

stitution at C_{12} and the double bond in the 9:11 position. The significantly further downfield position of this proton as compared to its 17-desoxy analog (IV), (Table 1) indicated the profound influence of the 17- α —OH group. The proximity of an α (axial) oriented hydrogen on C₁₂ to the C_{17 α}-OH group (1:3, diaxial type interaction) could well account for the observed differences.⁸ No such effect would be anticipated for the protons (C_6 —H, C_7 —H) of the alternative possibilities. The NMR investigations of Greene et $al.^9$ on a number of 9:11 dehydrotigogenins substituted on C_{12} with an OH, OAc or OMe group have demonstrated that in all those derivatives, in which the substituent is α (axial). both the vinylic proton, C₁₁-H, and the $C_{12}\beta$ —H proton appear as doublets with a coupling constant of ca. 6 c/s, while for the 12B-oxygenated (equatorial) derivatives both signals appear as broad singlets. The broadness of these signals in compound III results from the small coupling between the C_{11} and C_{12} protons as well as allylic and homoallylic coupling of the C_8 -proton (β , axial) with the vinylic and C_1 , \rightarrow H protons,^{7 α , 10} respectively (see below). Confirmatory evidence for the $9:11$ -dehydro-12 β -OMe assignment was forthcoming from the comparison of the chemical shift for the vinylic hydrogen (538 ppm), which corresponds to that reported for the C_{11} proton of 12 β -methoxy-9:11-dehydrotigogenin acetate (5:34)⁹ and not the 12 α -OMe analog (5:64).⁹ The vinylic proton for 7 α -methoxy-cholest-5-ene-3 β -diol¹¹ appears at 5.73 ppm ($J_{6, 78} = 5.5$ c/s). The C_{12} —H proton in both compounds, III and IV, we may note, appears significantly further downfield than that reported for 12β-methoxy-9-dehydrotigogenin acetate (3.42). We presume that the lactone moiety $(C_{18}-C_{20})$ is in part responsible for this.

17-Desoxy12~-methoxy-7,8-dihydro-22,25-oxidoholothurinogenin-3-acetate (IV) $C_{33}H_{50}O_6$, m.p. 281-282°, $[\alpha]_D^{25}$ -45°. In the UV, the compound showed no significant absorption above 215 mµ. IR (CS_2) absorption bands attributable to a γ -lactone (1761), acetyl (1730), side-chain ether (1132, 1107), and OCH₃ (1080 cm⁻¹), accounted for all the oxygens present in the molecule. No IR absorption band for an OH group was noted. Corresponding IR absorption bands (except for the OMe. group) are observed¹ in its derived holothurinogenin (II). The NMR (CDCl₃) spectrum showed the presence of one vinyl hydrogen (5.37, broad singlet; C_{11} —H); a broad singlet at 3.93 (C_{12} —H) partially overlapped by a just discernible triplet centered at 3.99 ($J = 7$ c/s, C₂₂—H). In benzene the signals for the C₁₂ and C₂₂ protons were clearly separated and appeared at 3.81 (broad singlet), and 4.07 (triplet, $J = 7$ c/s), respectively. The position and splitting of the triplet corresponded to that for the C₂₂—H proton of its derived holothurinogenin (II).¹ The 12 β -assignment for the OMe grouping was deduced following the same line of argument presented above for the 12ß-OMe substituted neo-holothurinogenin (III). Confirmation of these assignments was obtained in the 100 MC/S spectra and spin decoupling experiments. A clear separation of the C_{12} -H proton (3.88, broad singlet) and the C₂₂---H proton (3.95, triplet, $J = 7$ c/s) was achieved. Irradiation at 3.88 resulted in the transition of the broad signal for the vinylic hydrogen to a doublet $(J = ca$. 1.5 c/s); conversely, irradiation of the C_{11} proton (5.29) transformed the broad signal for the C₁₂ proton to a doublet ($J = ca$. 1 c/s). These decoupling experiments clearly demonstrated that allylic and homoallylic coupling of the C_8 proton with the C_{11} and C_{12} protons, respectively, was also present. Homoallylic coupling would only be anticipated, if both the C_{12} -H and C_8 -H bonds were approximately perpendicular to the plane of the 9:11 bond.¹⁰ An examination of Drieding models

demonstrated that this condition is fulfilled, when the protons on C_8 and C_{12} are β (axial) and α (axial), respectively.

12B-Methoxy-7,8-dihydroholothurinogenin-3-22, diacetate (VII) $C_{33}H_{54}O_8$, m.p. 310" (dec). Compound VII showed no absorption in the UV above 215 mu. IR (CHCl₃) absorption bands attributable to an OH (3604, 3500 bonded); γ -lactone (1753) ; 2-acetyls $(1720-1710)$ and OMe group (1086 cm^{-1}) accounted for the eight oxygens present in the molecule. The NMR $(CDCl₃)$ spectrum showed the presence of seven methyls $(0.88, 0.93, 1.25, 1.08, 1.55, 0.88)$ (2), $J = 6$ c/s); two acetyl methyls (2.08, 2.12); OMe (3.45); one vinyl hydrogen (5.37; C₁₁—H); one proton on C₁₂ bearing the OMe (4.13) ; and two protons on the C atoms bearing the acetyl groups (4.56, 5.08 ppm C_3 —H and C_{22} —H, respectively). The broad singlet nature of the protons on C_{11} and C_{12} established the C_{12} proton as axial (vide supra).

12B,25-Dimethoxy-7,8-dihydroholothurinogenin-3-acetate (VI) $C_{3.4}H_{5.4}O_7$, m.p. 272-273°, $\lceil \alpha \rceil_0^{25}$ - 51° (CHCl₃). The observed mol ion peak at *m/e* 574 as well as peaks at m/e 542, 510 (loss of one and two moles of methanol respectively) and m/e 559 (loss of a Me group) confirmed the molecular formula and the presence of two OMe groups. The compound showed no significant absorption in the UV above 215 m μ . The IR (CHCl₃) showed absorption bands for an OH (3600, 3480 bonded); γ -lactone (1750); acetyl (1720 broad), and OMe group (1090 cm⁻¹); v_{max} (KBr) 1743 (lactone) and 1722 (acetate). In CCl₄ the band for the acetate was split into two, 1728 and 1710 (small and sharp). This splitting of the acetyl in CCl_4 appears to be characteristic of the holothurinogenins belonging to the open side-chain series. We have noted the same behavior with holothurinogenin $(VIII)^5$, obtained from VI or holothurin A following strong acid treatment. The very limited solubility of compounds VI and VIII in CCI_4 suggests that in this solvent gels rather than true solutions are obtained. Hydrolysis of compound VI with alkali furnished the corresponding alcohol, $(R_1 =$ OH), whose IR showed only one band at 1751 cm⁻¹ (CHCl₃) and 1733 cm⁻¹ (KBr) attributable to the lactone. The NMR $(CDCl₃)$ spectrum showed the presence of one vinyl hydrogen (5.41; C₁₁—H); two OMe groups (3.45, 3.20; C₁₂ and C₂₅, respectively); and one proton (4.12, C_{12} —H) on the C atom bearing a OMe group (Table 1). The broad singlets, attributable to the protons on C_{11} and C_{12} , indicated that the C_{12} proton (*vide supra*) was axial. This was confirmed by spin decoupling experiments at 100 MC/S. Irradiation of the vinyl proton at 5.37 resulted in the collapse of the broad singlet at 4.12 to a doublet $(J = 1.5 \text{ c/s})$; alternatively irradiation at 4.12 ppm resulted in a sharpening of the signal at 5.41 . As with compound V (vide supra) the broadness of the signals for the protons on C_{11} and C_{12} arises from their coupling with each other and allylic and homoallylic coupling,¹⁰ respectively, with the proton (β , axial) on C_8 .

12β-Methoxy-7,8-dihydro-24,25-dehydroholothurinogenin-3-acetate (V) C₃₇H₅₀O₆; m.p. 245-247° (dec) $\lceil \alpha \rceil_{\text{D}}^{25}$ -53° (CHCl₃). IR (CCl₄), absorption bands, attributable to an OH (3600, 3500); y-lactone (1764); acetyl (1730), and OMe (1091 cm⁻¹) accounted for the six oxygens present in the molecule. The NMR spectrum (CDCl₃) showed seven methyls: five tertiary $(0.88, 0.93, 1.08, 1.27, 1.55)$ and two on a double bond $(1.67, 1.73)$; one OMe (3.45); two vinyl hydrogens (5.37, C₁₁—H; 5.16, C₂₄—H); and a proton on the C atom bearing the OMe group $(4.12, C_{12} - H)$. The chemical shift and broad singlet character of the signals attributable to the C_{11} and C_{12} protons corresponded in three solvents (CDC_1) , pyridine and benzene) with those observed

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 C_{23} =CH₂ isomer (15%). Recrystallization did not significantly alter the ratio of these two signals. In benzene and pyridine the latter signal was overlapped

by the C_3 – H prot

by the C₃-H proton.

19 ---- -.._ g with compounds VI, VII and clearly demonstrated that the OMe group was 128 (Table 1).

 12β -Methoxy-7,8-dihydroholothurinogenin-3-acetate (X) $C_{33}H_{52}O_6$; m.p. 263--267°. Compound X showed no absorption in the UV above 215 mu. The NMR spectrum showed the presence of seven methyls (0.88, 0.93, 1.08, 1.25, 1.53, 0.88(2), $J = 6$ c/s); one acetyl methyl (208); one OMe (3.45); one vinyl hydrogen (5.37, C_{11} —H); and one proton on the C atom bearing the OMe group $(4.11, C_{12} - H)$. The similar disposition of these signals as compared to griseogenin (IX) and compound V, coupled with the observation that compound X is only obtained from the hydrolysis of dihydroholothurin A, leaves little doubt as to the assigned structure.

The NMR spectra of the methoxylated neo-holothurinogenins have been examined in chloroform, benzene and pyridine. The results are summarized in Table 1. The various assignments follow from both the discussions given above, analogy with the holothurinogenins, and the reasonable assumption that an internally consistent picture should obtain for these closely related compounds. The Me groups on C_4 and C_{10} are essentially in the same environment in all these compounds and would have similar chemical shifts in all the solutions, as indicated in Table 1. As previously discussed¹ for the holothurinogenins I and II, the deshielding effect of the 17α —OH group (1,3-diaxial interaction) causes a downfield shift of the C_{14} Me in compound III as compared to compound IV. Thus, with the C_{21} assigned^{1, 12} the farthest downfield position and the C_{26} and C_{27} methyls equivalent (CDCl₃), the assignments for compounds III and IV are uniquely defined. The assignments of the Me groups of the "open side-chain" methoxylated neo-holothurinogenins, all of which contain a 17α —OH, follow by comparison with the compounds III and IV and the known structural interrelationships of the compounds within the group.

A number of interesting chemical shifts observed on solvent change (CDCl₃ \rightarrow C_6D_5N and/or C_6D_6) deserve comment. The chemical shifts for the methyls on C_4 are relatively little affected in the various solvents. Particularly noteworthy is the non-equivalence, observed in benzene and pyridine, of the C_{26} and C_{27} methyls of the .22,25-oxidoholothurinogenins (III and IV) as well as the relatively large downfield shift noted for one of these methyls. The C_{14} —Me is displaced significantly downfield in pyridine only and in only those compounds which contain the 17α —OH group, while the C_{10} and C_{21} methyls are displaced downfield in both benzene and pyridine. The magnitude of the shift for the C_{21} methyls in pyridine is again clearly dependent on whether the compound contains a 17α —OH group. In pyridine the C_{21} —Me group of compounds III, V and VI are displaced downfield 20 \pm 1 c/s as compared to CDCl₃; in compound IV (no 17 α —OH) the displacement is 8 c/s downfield. A similar comparison⁵ of the C_{21} Me in holothurinogenin II with the holothurinogenins I, VIII and IX showed the same effect of pyridine. No downfield shift was noted in compound II, while for compounds I, VIII and IX the signal appeared ca. 12 c/s downfield in pyridine as compared to chloroform. Clearly then the 12β — OMe group also contributes to the downfield shift of the C_{21} —Me. It appears reasonable to postulate that for pyridine to have such an effect, both, 17α —OH and 12 β —OMe must be close to the C₂₁—Me. In which case it may be argued that the C_{21} —Me is oriented β (behind the plane of the lactone ring) as in the lanosterol series.¹³ The α -orientation of the Me group places it perpendicular to the 17 α —OH and at much too great a distance from the 12β —OMe for pyridine molecules, presumably preferentially bound to these groups, to have any substantial magnetic effect (see following paper).

EXPERIMENTAL

The UV in 95% EtOH and IR spectra were recorded with a Beckman D.U. and Perkin-Elmer IR 421 spectrophctometer, respectively. NMR spectra were determined at 60 MC/S and 100 MC/S with Varian Associates spectrometers A-6OA and HA 100. Spin decoupling experiments were performed at 100 MC/S. The chemical shifts 8 are given in ppm relative to an internal TMS standard. M.ps were determined on a Fisher-Jones m.p. block. Alumina HA, prepared by the deactivation of Woehlm neutral alumina I with 4% by wt of a 10% AcOH soln, was employed for column chromatography. TLC (20 \times 20 cm glass plates) on silica gel G and Woehlm neutral alumina with water-sat BuOH-pyridine (3%) and benxene-EtOAc (9:1),respectively,asdevelopingagents.Tripledevelopments weregenerallyemployedanddistance travelled given in cm from origin. For detection of spots the plates were heated to 120", sprayed with a 1 :l mixture of anisaldehyde-AcOH-H₂SO₄ (1:50:0-5) and AcOH-HClO₄ (1:1) and reheated at 120° for 5 min.

Starting materials

Holothurin *A.* Holothuria (50 g) prepared, as previously described,' from Cuvier's giand of Acfinopyga agassizi was dissolved in boiling MeOH (31) and filtered from insoluble material $(3.5 g)$. The filtrate was boiled down to $\frac{1}{2}$ of its volume and allowed to stand overnight in the cold. The ppt (25 g) was recrystallized from MeOH containing H_2O (0-2-0-5%) and gave chromatographically homogeneous holothurin A (15 g) indistinguishable from material previously prepared via cholesterol precipitation,³ (α)²⁵ = -16° (H, O) , $R_f = 0.17$ (silica gel). Additional recrystallization did not alter its properties. The NMR and IR spectra will be discussed in a separate communication.¹⁴ Dihydroholothurin A was prepared¹⁴ by hydrogenation, at atmospheric pressure, of holothurin A in $H₂O$, employing PtO₂ as catalyst.

Methanol-HC1 *hydrolysis of holothurin A*

Compounds III, IV, V, VI and VII. Holothurin A (59 g) was suspended in MeOH-HCl(O2 N, 250 ml) (prepared by diluting cone HCI (4.24ml) to 25Oml with MeOH and the mixture stirred and heated at 50" for 76 hr. A clear soln was obtained in &5 hr. The hydrolysis was followed by removing aliquots for TLC (silica gel) and UV absorption. After 76 hr no glycoside remained and little conjugated diene (λ_{max} 244 m μ , e 1,500) was present in the hydrolysate. The product (1.9 g), $R_f = 0.77$ (silica gel), precipitated with H₂O (1.5 l.), was dried in vacuo over P₂O₅, dissolved in a mixture of pyridine (8 ml) and Ac₂O (4 ml)

Fraction	Eluent	Vol. (ml)	TLC (cm) ^{σ}	Material (mg)
A	Benzene-Skelly $B(1:3)$	1000	$18 - 5$	Oil (19)
B	Benzene-Skelly B $(1:1)$	235	14.0	IV(75)
C	Benzene-Skelly $B(1:1)$	250	$14, 7.5, (9)*(10), * (12)*$	$IV + III(110)$
D	Benzene-Skelly $B(1:1)$ Benzene-Skelly B (3:2)	650 600	$7.5'(9)^*, (10)^*, (12)^*$	III (397)
E	Benzene-Skelly B (3:2)	1100	7.0, (7.5)	V(86)
F	Benzene-Skelly B $(2:3:1)$	500	$7-0.4-0$	$VI + VII (32)$
G	Benzene-Skelly B $(2:3:1)$ Benzene-Skelly $B(6:1)$	1200 400	$4-0, (3-0)$	$VII + VI(105)$
H	Benzene Benzene-AcOEt (20:1)	700 150	$3-0, 4-0$	$VI + VII (58)$
	Benzene-AcOEt (20:1)	250	$3-0$	VI(439)
	Benzene-AcOEt (10:1)	200	3.0, 1.0	$VI + ?(93)$

TABLE 2

0 TLC on Woehlm neutral alumina; distance given in cm from origin following triple development at room temp; subsidiary spots in (); those starred ()* represent minor constituents not identilied. and the mixture allowed to stand at room temp for 3 days. The acetate mixture (1.95 g), precipitated with H₂O, was dried over P₂O₃ in vacuo, redissolved in benzene-Skelly B (10:1, 11 ml) and chromatographed on alumina HA (70 g; column (50 \times 2 cm) prepared in Skelly B). Elution was effected with Skelly Bbenzene and benzene-EtOAc mixtures. Fractions (25-30 ml) were collected and combined according to their TLC (alumina) and NMR spectra, as shown in Table 2.

17-Desoxy-12ß-methoxy-7,8-dihydro-22,25-oxidoholothurinogenin-3-acetate (IV)

Fraction B (75 mg) was recrystallized $(2 \times)$ from MeOH and gave compound IV, m.p. 281–282° (change in form at 265°); $[\alpha]_0^{2.5} = -45^\circ$ (CHCl₃), end absorption λ 213 mu (e 4900). For IR absorption in CS₂ see text; v_{max} (KBr) 1760 (lactone); 1720, 1248, 1030 (acetate); 1129, 1122 (side-chain ether); 1084 (OCH₃); 1061, 1051, 847cm-' (lactone and sidechain ether). No IR absorption in the OH region. For NMR spectra see Table 1. (Found: C, 73.54; H, 9.16. $C_{33}H_{50}O_6$ requires: C, 73.03; H, 9.29%).

17-Desoxy-22,25-oxidoholothurinogenin-3-acetate (II) from compound IV

Compound IV (10 mg) dissolved in a mixture of dioxan (2 ml) and HCI (3 N; 1.0 ml) was heated on a steambath for 3 hr. The product precipitated with H_2O was dried in vacuo over P_2O_5 and treated at 37° for 7 hr with a mixture of pyridine (1 ml) and $Ac₂O$ (0-5 ml). The acetylated product, obtained in the usual manner, was identical, NMR, TLC with an authentic sample' of compound II.

12B-Methoxy-7,8-dihydro-22,25-oxidoholothurinogenin-3-acetate (III)

Fraction D (390 mg) was recrystallized (3 X) from MeOH and gave compound III, m.p. $273-274^{\circ}$ (transition at ca. 250°), $[\alpha]_0^{25}$ - 60° (CHCl₃). For IR (CS₂) see text; v_{max} (KBr) 3568 (OH); 1752, (γ -lactone); 1722, 1252, 972 (acetate); 1088 (OCH₃) 1138 (side-chain ether); 1072, 1046, 841 cm⁻¹ (side-chain ether + lactone). For NMR spectra see Table 1. (Found: C, 70-98; H, 8.87. $C_{33}H_{50}O_7$ requires: C, 70-94; H, 9.02%).

22,25-Oxidoholothurinogenin-3-acetate (I)from compound III

Compound III (25 mg) was heated in dioxan-HCI and acetylated following the same procedure given above for compound IV. The final product proved identical in all respects, IR, NMR, m.p., TLC with an authentic sample' of compound I.

12B-Methoxy-7,8-dihydro-24,25-dehydroholothurinogenin-3-acetate (V)

Fraction E (86 mg) was recrystallized (2 x) from EtOAc and yielded compound V, m.p. 245-247°, $\lceil \alpha \rceil_0^{25}$ -53° (CHCl₃). For IR (CCl₄) see text; v_{max} (KBr) 3520 (OH); 1745, 1069, 1044 (γ -lactone); 1723, 1250, 979 (acetate); and 1089 cm^{-1} (OCH₃). For NMR spectra see Table 1. (Found: C, 72.55; H, 9.36. C₃₃H₅₀O₆ requires : C, 73.03 ; H, 9.29%).

12gMethoxy7,8-dihydroholothurinogenin-3,22-diacetate (VII)

Fraction G (105 mg) was recrystallized from EtOAc and gave compound VII, $m.p.$ 310 $^{\circ}$. For IR (CHCl₃) see text, for NMR s.c Table 1.

Griseogenin diacetate (IX) from compound VII

Compound VII (19 mg) was heated with dioxan-HCI and acetylated following the same procedure as given above. The product, recrystallized from MeOH, m.p. $256-257^{\circ}$, was identical in all respects, IR, UV, NMR, with an authentic sample⁶ kindly provided by Prof. C. Djerassi.

12~,25-Dimethoxy-7,8-dihydroholothurinogenin-3-acetafe (VI)

Fraction I (213 mg) was recrystallized (3 x from MeOH and gave compound VI, m.p. 272-273° (transition at ca. 260°), $[\alpha]_0^{25}$ - 52° (CHCl₃). For IR spectra in CHCl₃ and CCl₄ see text; v_{max} (KBr) 3519 (OH); 1743. 1067, 1042 (γ -lactone); 1722, 1250, 973 (acetate); 1088 cm⁻¹ (OCH₃); for NMR spectra see Table 1. (Found: C, 7@80; H, 9.51; moI.wt. 574 (mass spectroscopy). C34Hs407 requires: C, 71a5; II, 9.47%; moL wt. 574). The mass spectrum revealed the presence of two contaminants, m/e 572 and 568. The first probably a dehydro derivative of VI, the second presumably a fragment from a higher mol.wt. material, possibly the 12-acetate derivative of dehydro-VI (M_{600} —CH₃OH).

24,25-Dehydroholothurinogenin-3-acetate from compound VI

Compound VI was heated in dioxan-HCI and acetylated following the same procedures described

above. The product was chromatographed on alumina HA (2 g) and the material (10 mg), eluted with benzene (300 ml), was identical, NMR, UV, IR, with an authentic sample⁵ of compound VIII ($R_1 = OH$), prepared directly by strong acid hydrolysis of holothurin A.

Methanol-HCl hydrolysis of dihydroholothurin $A \rightarrow$ compounds III, IV, VII and X

Dihydroholothurin A (4 g) was suspended in MeOH-HCl (0-2 N, 200 ml) and the mixture stirred and heated at 50' for 66 hr. The course of the hydrolysis was' followed as described above for holothurin A. The product, precipitated by the addition of H₂O, contained ca. 15% of conjugated dienes ($\lambda_{\text{max}} = 244$ mµ, ϵ 3,000), was extracted with ether. The ether soluble residue (0.53 g) was dissolved in pyridine (6 ml) and Ac,O (3 ml) and the mixture allowed to stand one day at room temp. The acetylated product precipitated with H₂O was dried over P₂O₅ in vacuo, redissolved in benzene (3 ml) Skelly B (2) and chromatographed on a column of alumina IIA (20 g, prepared in Skelly B). Elution was effected with Skelly B-benzene-EtOAc mixtures. Fractions (25-30 ml) were collected and combined according to their TLC (alumina) and NMR spectra, as indicated in Table 3.

a TLC on Wcchlm neutral alumina at distance given in cm from origin following triple development; () indicate minor constituents, (\rightarrow unidentified material present in traces.

$17\text{-}Desoxy-12B\text{-}methoxy-7,8-dihydro-22,25-oxidoholothurinoaemin-3-acetate (IV)$

Fraction B was identified as compound IV by comparison, NMR, TLC, with the authentic sample prepared from holothurin A (see above),

22,25-Oxidoholothurinogenin-3-acetate (I)

Fraction C (33 mg) was recrystallized from MeOH and gave compound I (14 mg), identical in all respects, m.p., UV, IR, NMR, *R,* with an authentic sample,' prepared from holothurin A by strong acid hydrolysis.

12β-Methoxy-7,8-dihydroholothurinogenin-3-acetate (X)

12ß-Methoxy-7,8-dihydro-22,25-oxidoholothurinogenin-3-acetate (III). Fraction E (147 mg) was subjected to preparative TLC (Woehlm neutral alumina). Following triple development the material, migrating 90 and 8-0 (observed with UV), was extracted from the alumina with CHCl₃ and rechromatographed (TLC, triple development). Recrystallization of the slower moving compounent, 8-0, from MeOH gave compound X (17 mg), m.p. 263-267°. For NMR see text. (Found: C, 72.53; H, 9-68. $C_{33}H_{32}O_6$ requires: C, 72.76; $H, 9.62\%$).

Recrystallization of the faster moving component (9G) from MeOH gave compound III (20 mg) identical in all respects, m.p., NMR, with that prepared above.

12&Methoxy7,8-dihydroholothwinogmin-3,22-diocetote (VII)

Fraction G *(42* mg) was recrystallized from EtOAc and gave material identical in all respects, m.p., IR, NMR, with the sample (VII), prepared from holothurin A (see above).

Hydrolysis of dihydroholothurin A in dioxan-HCl (0-2 *N), followed by* MeOH-HCl (0-2 *N)*

Dihydroholothurin A (0.8 g), suspended in a mixture of dioxan (36 ml), H_2O (3.2 ml) and conc HCl (0.65 ml) was stirred and heated at 55" for 45 hr. A clear soln was obtained after 7 hr. The hydrolysis was followed by removing aliquots for TLC (silica gel) and UV absorption. After 45 hr no glycoside remained and ca. 20% of conjugated dienes (λ_{max} 244 m μ , ε 3,500) were present in the hydrolysate. The product (0-25 g, $R_f = 0.69$, silica gel) was precipitated with H₂O (250 ml) and dried in vacuo over P₂O₅ overnight. The NMR (CDCl₃) spectrum of the mixture showed broad singlets at δ 5.20 (C₁₁—H, vinyl proton), and 4.57 OH

 $\Delta_{\rm C}$ / Δ); overlapping multiplets, 4.3-3.6, representing the C_{22} proton of the oxido and OH substituted \angle \rightarrow

 \searrow \swarrow ^{OH} neo-holothurinogenins; a multiplet at 3.25 ($\overline{C_3}$ \prime) and no signal attributable to Me groups on a

double bonded C atom. The crude product (60 mg), dissolved in pyridine (2 ml) and $Ac₂O$ (1 ml), was allowed to stand at room temp for several hr. The acetate mixture (40 mg), precipitated with H_2O , was

H

OAc dried in vacuo over P₂O₅. The NMR (CDCI₃) spectrum showed broad singlets at 5.75 ($\overrightarrow{C_{12}}$) and ' 'H 'H 'H

OAc 5.12 (C₁₁-H), a multiplet at 4.57 ($\sum_{n=1}^{\infty}$ $\overline{}$); a broad triplet at 4.12 (C_{22} —H of oxido ring), and singlets H

at 2-08 (C₃ and C₁₂ acetyls), and 2·12 (C₂₂ acetyl). A multiplet corresponding to the C₂₂ was covered

OAc

by the broad singlet at 5.12

The aforementioned aglycone mixture (5Omg). obtained from dioxan-HCI hydrolysis (see above), suspended in methanol (5 ml) and conc. HCl (0.08 ml), was stirred and heated at 53° for 24 hr. The product (45 mg), precipitated with H₂O, was dried in vacuo over P_2O_3 . The NMR (CDCl₃) spectrum, which OMe

showed broad singlets at δ 5.37 (C₁₁—H, vinyl proton) and 4.12 (ζ \prime \rightarrow) (the latter signal covers

the triplet for the C₂₂—H proton of the oxido ring), and a singlet at 3.43 (C₁₂—OCH₃) was identical with the NMR spectrum of the mixture obtained directly from dihydroholothurin A on hydrolysis in MeGH- $HC1 (0.2N)$ at 55 $^{\circ}$ for 24 hr.

Acknowledgements-The authors wish *to* express their thanks to the National Science Foundation (Grant GB-6645), Office of Naval Research (Contract 107 543), and the National Institutes of Health (Grant AM 06180) for their generous support. We gratefully acknowledge the help of Prof. C. Djerassi, who so kindly furnished the mass spectroscopy measurements, and Dr. W. H. McFadden and Mr. M. Jacobs of IFF Inc., Union Beach, N.J., for the 100 MC spin decoupling experiments. We also wish to express our appreciation to the Lerner Marine Laboratory, Bimini, Bahamas, who made their facilities available for the collection of Actinopyga agassixi.

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